



Albert Einstein College of Medicine
OF YESHIVA UNIVERSITY

An Introduction to the shRNA Core Facility

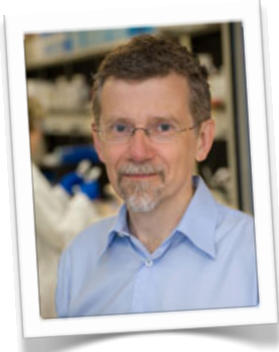
John Reidhaar-Olson, Ph.D.

January 25, 2012

RNAi User Group Meeting

shRNA Core Facility

Price 268 • www.einstein.yu.edu/sr/shRNA • shRNA@einstein.yu.edu



John Reidhaar-Olson, Ph.D.

- Ph.D. in Biochemistry from MIT
- Post-doc at UCSF
- >15 years in biotech and pharma, most recently at Roche
- Expertise in RNAi-based screening, RNA therapeutics, genomics



Deborah Smith, Ph.D.

- Ph.D. in Molecular Biology from NYU
- Post-doc at Sloan-Kettering
- Worked at high-throughput screening centers at Columbia and Yale
- Expertise in assay development, screening, and RNAi

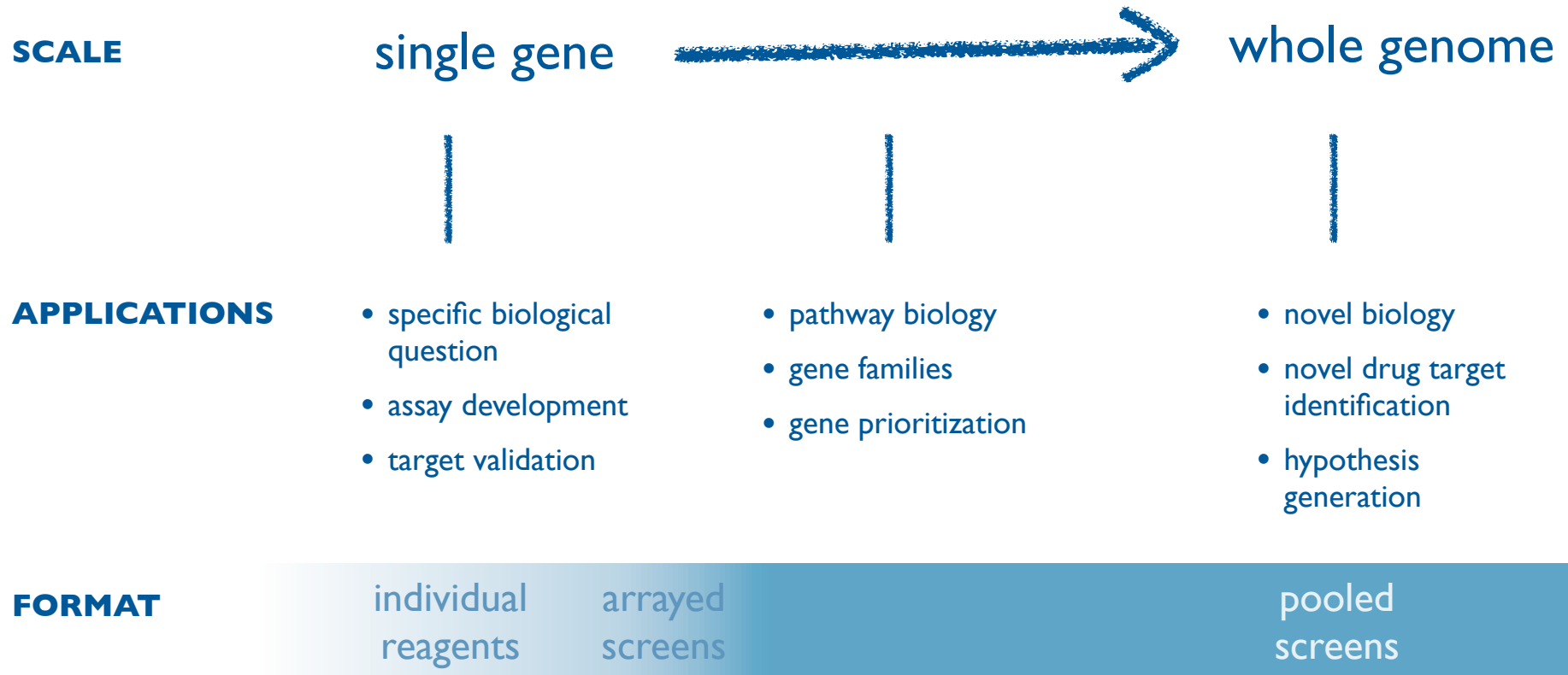


Deyan Tong, M.S.

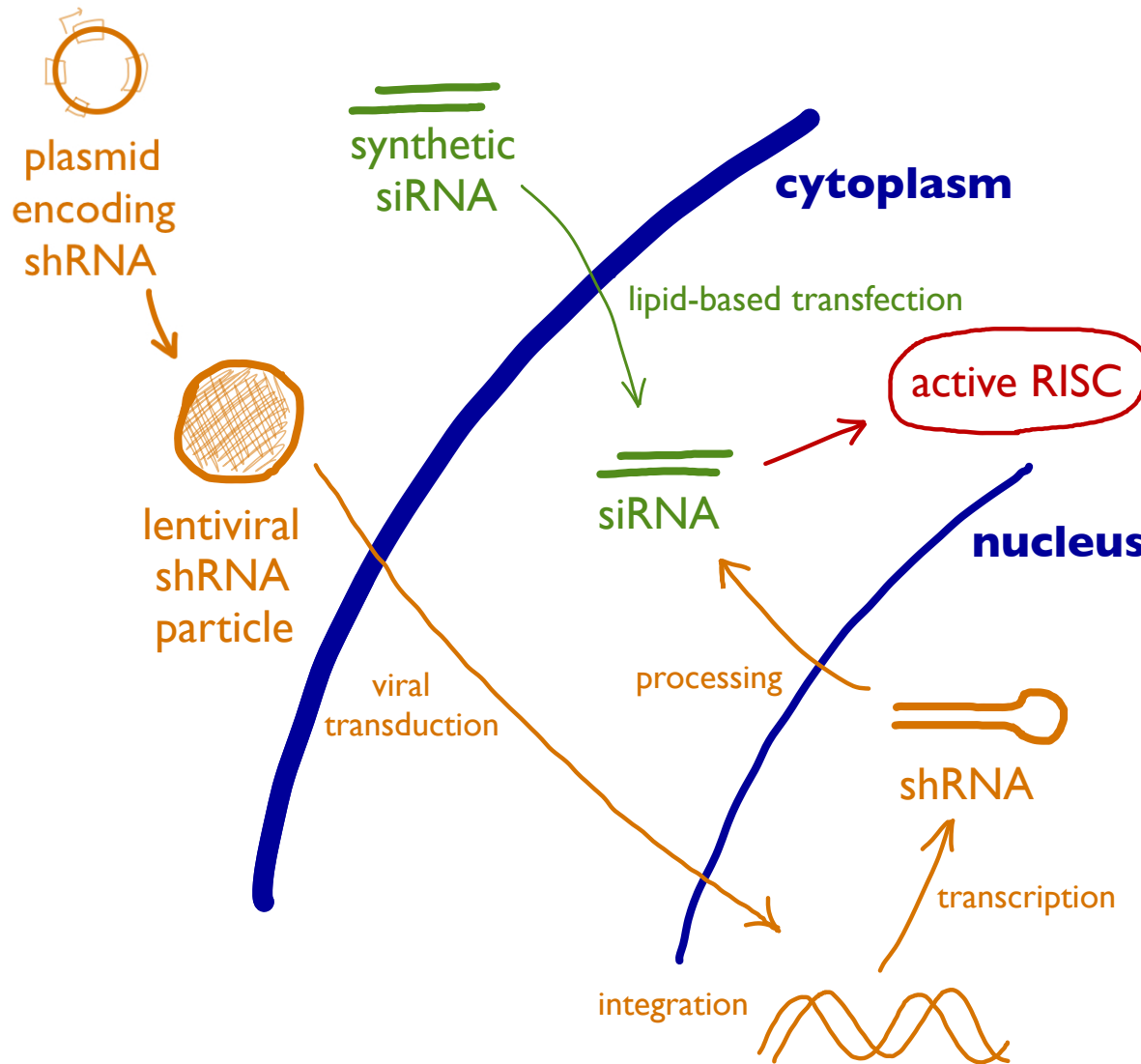
- Master's degree from University of Wisconsin–Madison
- Experience with lentivirus cloning, preparation and transduction

Applications

The shRNA Core Facility is dedicated to providing researchers with access to reagents, expertise, and infrastructure to enable RNAi-based loss-of-function studies at scales ranging from individual genes to genome scale.



siRNA and shRNA as means to silence gene expression



siRNA

- straightforward process
- highly effective knockdown
- effect is transient
- some cell types cannot be transfected
- limited to arrayed screens

shRNA

- more involved process
- broadly applicable to most cell types
- stable knockdown
- less off-target silencing
- allows for marker to follow transduction
- compatible with both arrayed screens and pooled selections

shRNA libraries at Einstein

Arrayed libraries

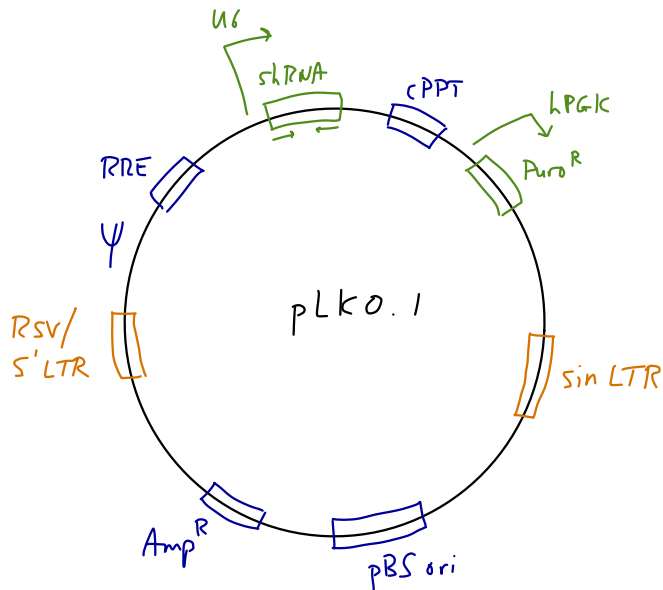
- Human
 - ▶ The RNAi Consortium (TRC) genome-wide shRNA collection
 - ▶ GIPZ lentiviral shRNAmir library (release 6.1–6.28)
 - ▶ Human Precision LentiORF Library
- Mouse
 - ▶ TRC genome-wide shRNA collection
 - ▶ GIPZ lentiviral shRNAmir library (release 7.1–7.17)



Pooled libraries

- Human
 - ▶ Decode RNAi-GIPZ whole genome pooled screening library
 - ▶ Decipher Human shRNA Libraries, modules 1, 2, and 3

TRC libraries



Human

- 18,000 genes
- ~4 shRNA clones per gene

Mouse

- 15,000 genes
- ~4–5 shRNA clones per gene

Rat (from human and mouse libraries)

- 10,000 genes
- ~2 shRNA clones per gene

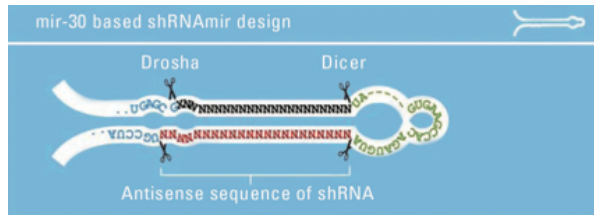
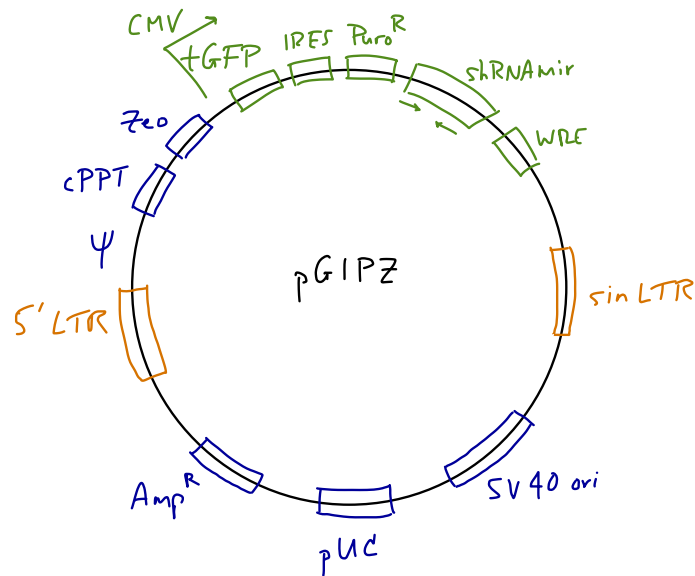
Developed by The RNAi Consortium based at the Broad Institute

shRNA	shRNA hairpin
U6	RNA Pol III promoter
Puro ^R	Puromycin resistance for selection in mammalian cells

Root, D. E., Hacohen, N., Hahn, W. C., Lander, E. S. and Sabatini, D. M. (2006) Genome-scale loss-of-function screening with a lentiviral library. *Nature Methods* 3:715–719.

Moffat J, Grueneberg DA, Yang X, Kim SY, Kloepfer AM, Hinkle G, Piqani B, Eisenhaure TM, Luo B, Grenier JK, Carpenter AE, Foo SY, Stewart SA, Stockwell BR, Hacohen N, Hahn WC, Lander ES, Sabatini DM, Root DE. (2006) A lentiviral RNAi library for human and mouse genes applied to an arrayed viral high-content screen. *Cell* 124:1283.

GIPZ library



Human

- 18,000 genes
- ~5 shRNA clones per gene

Mouse

- 15,800 genes
- ~3 shRNA clones per gene

Rat (from human and mouse libraries)

- 10,000 genes
- ~2 shRNA clones per gene

Developed by Greg Hannon and Steven Elledge

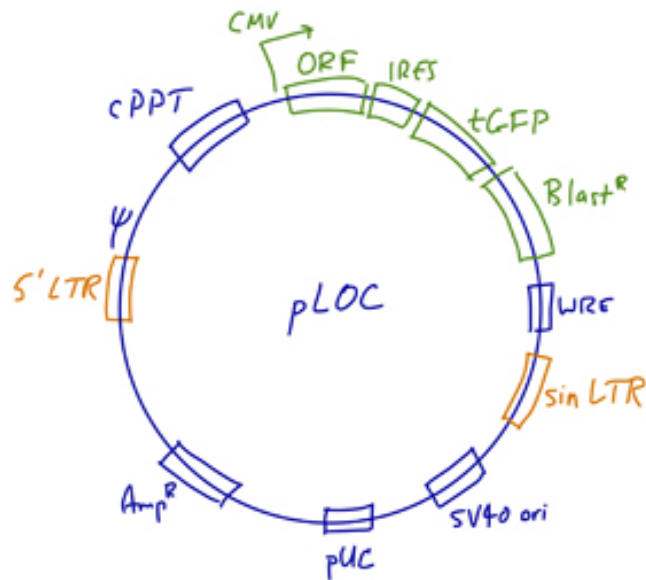
shRNAmir shRNA hairpin with mir-30 loop and context sequences to improve Drosha processing

tGFP turbo-GFP marker to track expression in transduced cells

CMV RNA Pol II promoter

Puro^R Puromycin resistance for selection in mammalian cells

LentiORF library



Human

- Currently 6,192 genes
- One ORF clones per gene

Based on content from the sequence-validated ORFeome Collaboration collection

Gateway vector format

ORF Open reading frame
 CMV RNA Pol II promoter
 tGFP turboGFP marker
 Blast^R Blasticidin resistance for selection in mammalian cells

Clones do not contain the naturally-occurring stop codon, to facilitate cloning of fusion tags

Source (ORF) Clone Library Information

ORF and ORF vector sequence transferred with ORF during Gateway cloning into LentiORF vector (Variable by source clone)

LentiORF Vector Sequence

Clone Source	Donor vector	Entry vector	ORF-	Vector sequence (from ORF clone)	att site	Vector	Fusion tag cloning	Synthetic STOP
DFCI-CCSB	pDONR223	pENTR223	ORF-	TGC	CCA ACT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				C	P T F L Y	K V V	A S	* *
DFCI-CCSB	pDONR223	pENTR223	ORF-	TAC	CCA ACT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				Y	P T F L Y	K V V	A S	* *
DFCI-CCSB (ORFeome 5)	pDONR223	pENTR223	ORF-	TTG	CCA ACT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				L	P T F L Y	K V V	A S	* *
DFCI-CCSB (ORFeome 1&3)	pDONR223	pENTR223	ORF-		CCA ACT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
					P T F L Y	K V V	A S	* *
WTSI	pDONR223	pENTR223	ORF-		CCA ACT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
					P T F L Y	K V V	A S	* *
HIP	pDONR221§	pENTR221	ORF-	TTG GAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				L D	P A F L Y	K V V	A S	* *
Kazusa	pDONR201§	pENTR201	ORF-	TAC GTA GAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				Y V D	P A F L Y	K V V	A S	* *
DKFZ	pDONR221§	pENTR221 (KnR)	ORF-	TGG ATC CAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				W I H	P A F L Y	K V V	A S	* *
DKFZ	pDONR221§	pENTR221 (KnR)	ORF-	AAG CTT GAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				K L D	P A F L Y	K V V	A S	* *
DKFZ	pDONR221§	pENTR221 (KnR)	ORF-	TGT ATT CAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				C I H	P A F L Y	K V V	A S	* *
DKFZ	pDONR221§	pENTR221 (KnR)	ORF-	GGC GAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				G D	P A F L Y	K V V	A S	* *
DKFZ	pDONR201§	pENTR201 (KnR)	ORF-	GGC GAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				G D	P A F L Y	K V V	A S	* *
MGC Synthetic	pDONR223.1	pENTR223.1	ORF-	TCA GGC CTC ATG GGC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				S G L M G	P A F L Y	K V V	A S	* *
DKFZ	None	pENTR/D-(KnR)	ORF-	AGG GGT GGG CGC GCC GAC	CCA GCT TTC TTG TAC	AAA GTG GTT	GCT AGC	TAA TGA
				K G G R A D	P A F L Y	K V V	A S	* *

shRNA Facility services

CURRENT

Reagent distribution from our shRNA and LentiORF libraries

- Glycerol stocks
- Plasmid DNA
- Lentivirus
 - ▶ Viral supernatant ($>10^6$ TU/ml)
 - ▶ High-titer viral stock ($>10^8$ TU/ml)
- Assessment of knockdown or overexpression by branched DNA assay

RNAi expertise

- Experimental design and data analysis



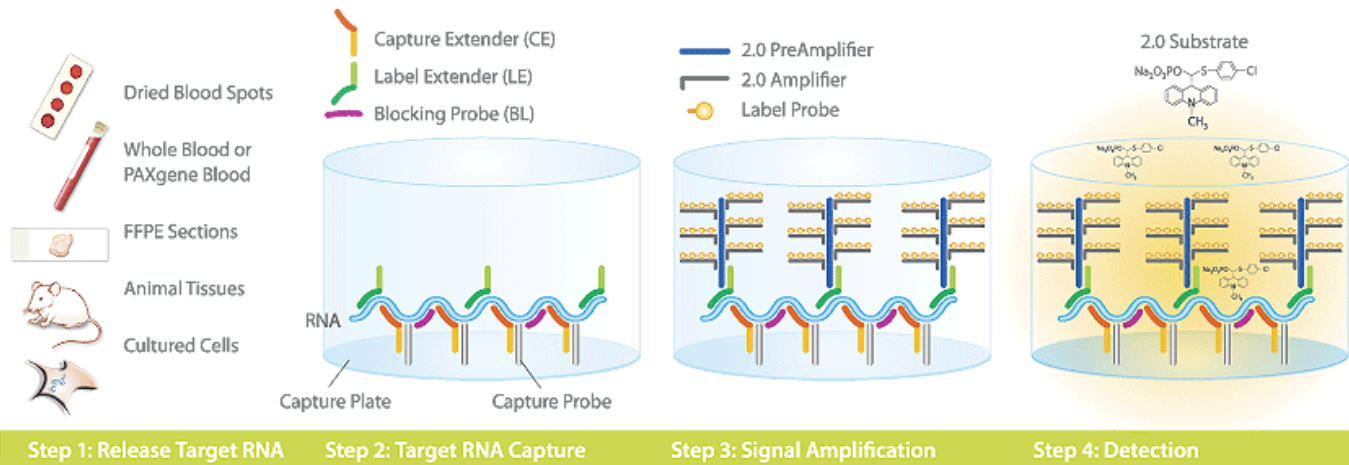
FUTURE

Screening services

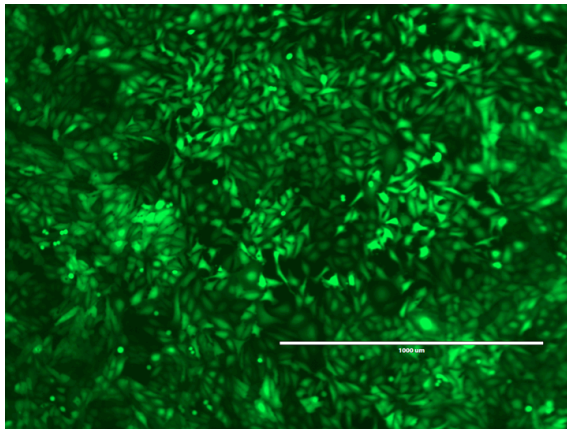
- Arrayed shRNA sets, provided as lentiviral preps
 - ▶ Pre-defined shRNA sets (e.g. to common pathways)
 - ▶ Custom sets
- Pooled screens
 - ▶ Genome-wide or large subset screens using pooled libraries
 - ▶ Custom pooled screening sets
- Assay support for screens
 - ▶ Plate reader and high content assays
- Hit identification assistance for pooled screens

QuantiGene branched DNA assay from Panomics

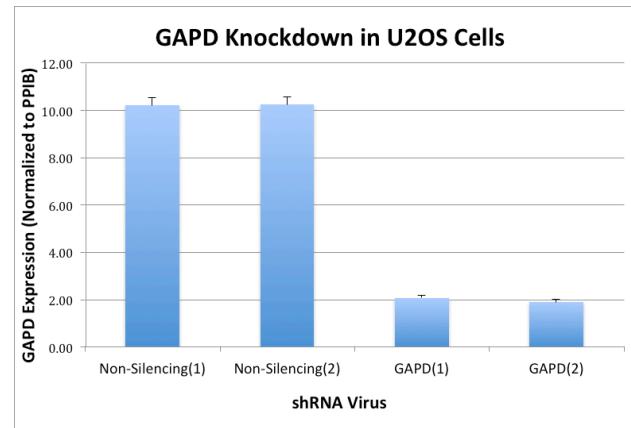
For quantitation of mRNA following shRNA-mediated knockdown



panomics.com



U2OS cells transduced with lentivirus containing the pGIPZ vector encoding GFP and shRNA targeting GAPDH



shRNA-mediated GAPDH knockdown in U2OS cells. Error bars represent standard deviations of technical triplicates.

Ordering shRNA reagents

Contact us (shRNA@einstein.yu.edu) with a list of genes you are interested in targeting

- Species (human, mouse, or rat)
- Library (GIPZ or TRC)
- Gene — provide gene symbol (e.g. GAPDH or ACTB) and gene ID (e.g. 2597 or 60)

We will send you a list of available clones and an order form

Einstein shRNA Core Facility

Click Oligo ID for more information from TRC or Open Biosystems websites. Click accession numbers to open RefSeq records.
An "S" in the "Alignment" column indicates a possible off-target effect arising from the sense strand.
Do not delete hidden columns

Human clones													
Gene Symbol	Gene ID	Catalog Number	Oligo ID	Production Note	Sense Sequence	Top BLAST hit to desired mRNA	Alignment	Target site	Top BLAST hit to other mRNA	Alignment	Top BLAST hit to other RNA (e.g. ncRNA)	Alignment	Library
ACTB	60	RHS4430-100994873	V3LHS_358210		AGCACAAATGAAGATCAAGA	NM_001101.3	19/19	1051 - 1069	NM_001199954.1	18/19	NR_037688.1	18/19	Hs_GIPZ
ACTB	60	RHS4430-98854611	V2LHS_233110		CATTGCTTTCGTGTAATT	NM_001101.3	19/19	1578 - 1595	NM_005708.3	16/19	XR_108735.2	19/19 S	Hs_GIPZ
ACTB	60	RHS4430-99139928	V2LHS_94242		GAAATCGTGCCTGACATTA	NM_001101.3	19/19	703 - 721	NM_001099771.2	17/19	XM_929706.4	17/19	Hs_GIPZ
ACTB	60	RHS4430-100991313	V3LHS_358211		TGGGACGACATGGAGAAAA	NM_001101.3	19/19	319 - 337	NM_001199954.1	17/19	NR_037688.1	17/19	Hs_GIPZ
ACTB	60	RHS4430-100995794	V3LHS_358208		CCCAGCACAAATGAAGATCA	NM_001101.3	19/19	1048 - 1065	NM_024068.3	16/19 S	NR_037688.1	18/19	Hs_GIPZ
ACTB	60	RHS4430-98521746	V2LHS_262423		CAAATATGAGATGCATTGT	NM_001101.3	18/19	1465 - 1483	NM_006197.3	15/19	XR_108735.2	18/19 S	Hs_GIPZ
ACTB	60	RHS3979-9596821	TRCN0000029413		CGAAACTACCTTCAACTCCAT	NM_001101.3	21/21	909 - 929	NM_001099771.2	20/21	NR_004845.1	21/21	Hs_TRC
ACTB	60	RHS3979-9596818	TRCN0000029410		CGTGCCTGACATTAAGGAGAA	NM_001101.3	21/21	708 - 728	NM_017450.2	17/21	NR_045673.1	15/21	Hs_TRC
ACTB	60	RHS3979-9596820	TRCN0000029412		GTTGCTATCCAGGCTGTGCTA	NM_001101.3	21/21	484 - 504	NM_003511.2	15/21	NR_002929.2	14/21	Hs_TRC

Available controls

TRC libraries

- Empty pLKO.1 vector (no shRNA insert)
- pLKO.1 clone targeting eGFP (can function as positive control or as non-targeting control)

GIPZ libraries

- Non-silencing control
- GAPDH shRNAmir (positive control)

LentiORF library

- pLOC clone encoding RFP (in addition to the GFP reporter)

Suggested practices

Test several shRNAs for each gene of interest

- Aim for at least two giving the phenotype of interest
- Make sure the functional shRNAs are independent (i.e. non-sequence-overlapping)

Include appropriate negative controls

- Non-targeting shRNA
- Non-transduced cells

Confirm knockdown at the protein or mRNA level

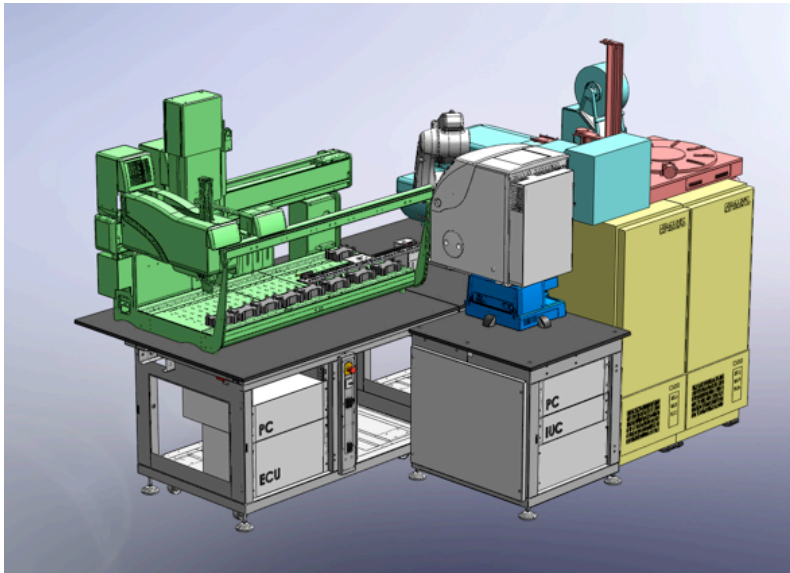
- Western blot, qRT-PCR, branched DNA

Confirm knockdown and phenotype in additional cell types, when possible

Consider rescue experiments when feasible

- Use an shRNA-resistant lentiORF to test for reversal of phenotype

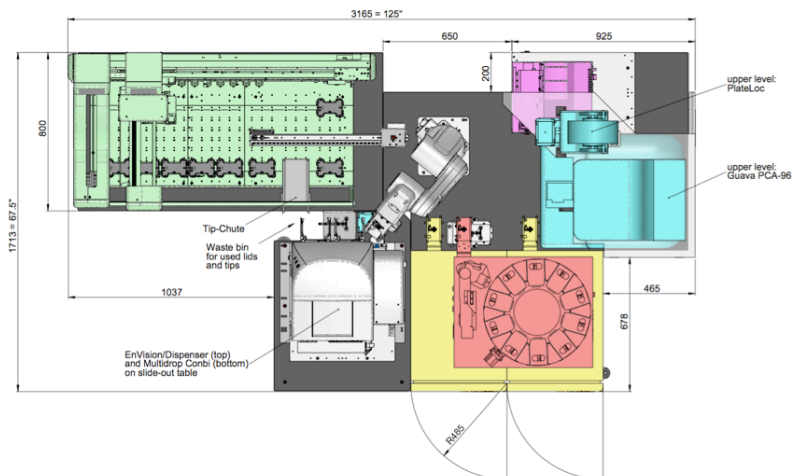
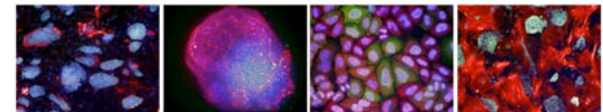
shRNA Core Facility equipment



EnVision
plate reader



High-content
imaging system
(later in 2012)



Robotics system with biosafety
enclosure

Contact

john.olson@einstein.yu.edu or shRNA@einstein.yu.edu

718-678-1195 • Price 275 (office), 268 (lab)

www.einstein.yu.edu/sr/shRNA

The screenshot shows the website for the shRNA Core Facility at the Albert Einstein College of Medicine. The header includes the Einstein logo and navigation links for 'About Einstein', 'Departments & Centers', 'Clinical Partners', 'Admissions', 'Research', and 'Library'. The main content area features a 'Shared Facilities' sidebar with links to 'home', 'libraries', 'services', 'ordering', 'instrumentation', 'resources', 'rna user group', 'faq', 'staff', and 'contact'. The main text area is titled 'shRNA Core Facility' and 'SHRNA', with a welcome message and a description of the facility's services. A green fluorescence microscopy image is displayed on the right side of the text. The footer contains the slogan 'Science at the heart of medicine' and various footer links and contact information.

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Shared Facilities | **shRNA Core Facility**

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SHRNA
Welcome to the Albert Einstein College of Medicine shRNA Core Facility
The Albert Einstein College of Medicine shRNA Core Facility is a state-of-the-art RNAi core facility dedicated to providing researchers access to reagents and infrastructure that enable loss-of-function studies at scales ranging from individual gene to genome scale. We provide RNAi reagents, shRNA-based screening services, and RNAi expertise to the Einstein research community. In addition, we offer some services to members of the New York Structural Biology Center as well. The Facility houses four genome-wide [shRNA libraries](#), from which we provide individual shRNA clones, shRNA sets for arrayed screens, and shRNA pools for selection-based screening. The Facility is [located](#) in the Price Center on the Einstein campus in the Bronx, and is affiliated with the [Albert Einstein Cancer Center](#).



Science at the heart of medicine

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